

THE UNIVERSITY of TEXAS

HEALTH SCIENCE CENTER AT HOUSTON SCHOOL of HEALTH INFORMATION SCIENCES

# Molecular Dynamics Simulation: Overview

For students of HI 6327 "Biomolecular Modeling"

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http://biomachina.org/courses/modeling/04.html

#### Protein (Biomolecular) Folding



native structure N



exploring the folding free energy (entropy + enthalpy) landscape

### Use of Molecular Dynamics Simulation

- •structure prediction / modeling
- •flexibility
- •solvent effects
- •chemical reactions (with QM)
- •conformational changes, allosteric mechanisms
- •thermodynamics (free energy changes, binding)
- •NMR/crystallography (refinement)
- •Electron microscopy (flexible fitting)

### Atomic Detail Computer Simulation



## Steric Energy

•A molecule can possess different kinds of energy such as bonded, nonbonded (both *enthalpic* contributions), and thermal energy (*entropy*).

•Molecular mechanics calculates the enthalpic or *steric* energy of a molecule - the energy due to the geometry or conformation of a molecule.

•Energy is *minimized* in nature, and the native conformation of a molecule that is favored is the lowest steric energy conformation.

•Studies of the conformation of proteins are difficult and therefore interesting, because their size makes many different conformations possible.

•In MD, steric energy allows the integration of the equations of motion following Newton's law.

## Steric Energy

Molecular mechanics *assumes* the steric energy of a molecule to arise from a few, specific interactions within a molecule. These interactions include the stretching or compressing of bonds beyond their equilibrium lengths and angles, torsional effects of twisting about single bonds, the van der Waals (vdW) attractions or repulsions of atoms that come close together, and the electrostatic interactions (qq) between partial charges in a molecule. To quantify the contribution of each, these interactions can be modeled by a potential function that gives the energy of the interaction as a function of distance, angle, or charge.

The total steric energy of a molecule can be written as a sum of the energies of the interactions:

$$E_{\text{steric energy}} = E_{\text{str}} + E_{\text{bend}} + E_{\text{improper}} + E_{\text{tor}} + E_{\text{vdW}} + E_{qq}$$

#### **Bonded Interactions: Stretching**

E<sub>str</sub> represents the energy required to stretch or compress a covalent bond:



A bond can be thought of as a spring having its own equilibrium length, r<sub>o</sub>, and the energy required to stretch or compress it can be approximated by the Hookean potential for an ideal spring:

$$E_{str} = \frac{1}{2} k_{s,ij} (r_{ij} - r_o)^2$$

#### **Bonded Interactions: Bending**

 $E_{bend}$  is the energy required to bend a bond from its equilibrium angle,  $\theta_o$ :



Again this system can be modeled by a spring, and the energy is given by the Hookean potential with respect to angle:

$$E_{bend} = \frac{1}{2} k_{b,ijk} (\theta_{ijk} - \theta_o)^2$$

### **Bonded Interactions: Improper Torsion**

 $E_{improper}$  is the energy required to deform a planar group of atoms from its equilibrium angle,  $\omega_0$ , usually equal to zero:



Again this system can be modeled by a spring, and the energy is given by the Hookean potential with respect to planar angle:

$$E_{improper} = \frac{1}{2} k_{o,ijkl} (\omega_{ijkl} - \omega_o)^2$$

#### **Bonded Interactions: Torsion**

E<sub>tor</sub> is the energy of torsion needed to rotate about bonds:



Torsional interactions are modeled by the potential:

 $\mathsf{E}_{\mathsf{tor}} = \frac{1}{2} \, k_{\mathsf{tor},1} \, (1 - \cos \phi) + \frac{1}{2} \, k_{\mathsf{tor},2} \, (1 - \cos 2 \phi) + \frac{1}{2} \, k_{\mathsf{tor},3} \, (1 - \cos 3 \phi)$ asymmetry (butane)
2-fold groups e.g. COO- standard tetrahedral torsions

#### Non-Bonded Interactions: van der Waals

 $E_{vdW}$  is the steric exclusion and long-range attraction energy (QM origins):



Two frequently used formulas:

$$\mathsf{E}_{VDW}(R) = \frac{A}{R^{12}} - \frac{B}{R^6} \qquad \qquad \mathsf{E}_{VDW}(R) = 4\varepsilon \big( \big(\frac{\sigma}{R}\big)^{12} - \big(\frac{\sigma}{R}\big)^6 \big)$$

#### Non-Bonded Interactions: Coulomb

 $E_{qq}$  is the Coulomb potential function for electrostatic interactions of charges:



The Q<sub>i</sub> and Q<sub>j</sub> are the partial atomic charges for atoms i and j separated by a distance  $r_{ij}$ .  $\epsilon$  is the relative dielectric constant. For gas phase calculations  $\epsilon$  is normally set to 1. Larger values of  $\epsilon$  are used to approximate the dielectric effect of intervening solute ( $\epsilon$ ~60-80) or solvent atoms in solution. k is a units conversion constant; for kcal/mol, k=2086.4.

#### Newton's Law

$$\mathsf{E}_{\mathsf{steric energy}} = \mathsf{E}_{\mathsf{str}} + \mathsf{E}_{\mathsf{bend}} + \mathsf{E}_{\mathsf{improper}} + \mathsf{E}_{\mathsf{tor}} + \mathsf{E}_{\mathsf{vdW}} + \mathsf{E}_{\mathsf{qq}}$$

Potential Function  $\rightarrow$  Force



Newton's Law:

 $F_i = m_i a_i$ 

#### Verlet's Numeric Integration Method

Taylor expansion:

$$r(t + \delta t) = r(t) + v(t)\delta t + \frac{1}{2}a(t)\delta t^{2}$$

$$r(t - \delta t) = r(t) - v(t)\delta t + \frac{1}{2}a(t)\delta t^{2}$$

$$r(t + \delta t) = 2r(t) - r(t - \delta t) + a(t)\delta t^{2}$$

#### Verlet's Method

#### **Timescale Limitations**



•Protein Folding milliseconds/seconds (10<sup>-3</sup>-1s) •Ligand Binding micro/milliseconds ( $10^{-6}$ - $10^{-3}$  s) •Enzyme catalysis micro/milliseconds (10<sup>-6</sup>-10<sup>-3</sup> s) •Conformational transitions pico/nanoseconds  $(10^{-12}-10^{-9} \text{ s})$ •Collective vibrations -1 picosecond ( $10^{-12}$  s) •Bond vibrations -1 femtosecond ( $10^{-15}$  s)

### **Timescale Limitations**



#### **Molecular dynamics:**

Integration timestep - 1 fs, set by fastest varying force.

Accessible timescale: about 10 nanoseconds.

## **Cutting Corners**

- Nonbonded interactions require order N<sup>2</sup> computer time!
  - Truncating at  $R_{cutoff}$  reduces this to order N  $R_{cutoff}^{3}$
  - Particle mesh Ewald (PME) method adds long range electrostatics at order N log N, only minor cost compared to cutoff calculation.
     PME, Schlick 9.4
- Can we extend the timestep, and do this work fewer times?
  - Bonds to hydrogen atoms, which require a 1fs timestep, can be held at their equilibrium lengths, allowing 2fs steps. SHAKE, Schlick 12.5
  - Long range electrostatics forces vary slowly, and may be evaluated less often, such as on every second or third step. MTS, Schlick 13.3

### Input Files: PDB

- Simulations start with atomic structures from the Protein Data Bank, in the standard PDB file format.
- PDB files contain standard records for species, tissue, authorship, citations, sequence, secondary structure, etc.
- We only care about the atom records...
  - atom name (N, C, CA)
  - residue name (ALA, HIS)
  - residue id (integer)
  - coordinates (x, y, z)
  - occupancy (0.0 to 1.0)
  - temp. factor (a.k.a. beta)
  - segment id (6PTI)
- No hydrogen atoms! (We must add them ourselves.)



#### Input Files: PDB

Go to http://www.rcsb.org/pdb/ and explain file content of example PDF file, e.g. PDB entry 1ATN

### Input Files: PSF

- Every atom in the simulation is listed.
- Provides all static atom-specific values:
  - atom name (N, C, CA)
  - atom type (NH1, C, CT1)
  - residue name (ALA, HIS)
  - residue id (integer)
  - segment id (6PTI)
  - atomic mass (in atomic mass units)
  - partial charge (in electronic charge units)
- What is not in the PSF file?
  - coordinates (dynamic data, initially read from PDB file)
  - velocities (dynamic data, initially from Boltzmann distribution)
  - force field parameters (non-specific, used for many molecules)

## Input Files: PSF



### Input Files: PSF

PSF example: download http://biomachina.org/courses/modeling/download/example.psf

### Input Files: Topology Files

blueprints for building a PSF file

- For every type of residue known:
  - atom name, type, mass, and charge
  - bonds within the residue
  - bonds to other residues
  - any planar impropers (rare)
- Additional "patches" for:
  - terminating protein segments
  - joining protein segments
  - modifying protonation states
  - adding disulphide bonds
  - deoxygenating nucleic acids



### Input Files: Topology Files

blueprints for building a PSF file

Example: download http://biomachina.org/courses/modeling/download/topallh22x.pro

#### Input Files: Parameter Files

defining the MM energy terms

- Equilibrium value and spring constant for
  - every pair of atom types that can form and bond
  - every triple of atom types that can form an angle
  - every quad of atom types that can form a dihedral or improper (many wildcard cases)
- vdW radius and well depth for every atom type
  - actually need these for every pair of atoms types!
  - pair radius calculated from arithmetic mean
  - pair well depth calculated from geometric mean
- Closely tied to matching topology file!

#### Input Files: Parameter Files

defining the MM energy terms

Example: download http://biomachina.org/courses/modeling/download/parallh22x.pro

### Setting up a MD Simulation

example: ubiquitin

- Obtain file from PDB (1ubq);
- Add missing hydrogen atoms;
- Determine protonation state of HIS residues;
- Add a water box;
- Trim the water box down to a sphere.





## Production Run Protocol

#### equilibrium properties



energies: kinetic and potential



exploring conformations



exploring conformations



exploring conformations

Actin SMD movies, see URL

http://www.biomachina.org/research/projects/actin

Go To:

2. Actin Phosphate Release – The Movie

### Further Reading

WWW:

http://cmm.info.nih.gov/intro\_simulation

Textbooks:

Schlick, Chapters 8, 9, 12, 13

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